



Bio-preservation of Foods: A Review

Vaishali^{1*}, Punit Jhandai¹, Vijay J. Jadhav¹ and Renu Gupta¹

¹*Department of Veterinary Public Health and Epidemiology, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana, India.*

Authors' contributions

This work was carried out in collaboration among all authors. Author VJJ conceptualized the review article. Author RG designed the framework of article. Authors Vaishali and PJ collected the references and prepared the draft. Authors VJJ and RG extensively edited the article to its final form. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/EJNFS/2019/v11i430159

Editor(s):

(1) Dr. Dan-Cristian Vodnar, Faculty of Food Science and Technology, University of Agricultural Science and Veterinary Medicine Cluj-Napoca, Cluj-Napoca, Romania.

Reviewers:

(1) Davies, Cristina Verónica, Universidad Nacional de Entre Ríos, Argentina.

(2) Rosendo Balois Morales, Universidad Autonoma de Nayarit, Mexico.

(3) Balogun Olalekan Blessing, Nigeria.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/53215>

Review Article

Received 20 October 2019

Accepted 23 December 2019

Published 30 December 2019

ABSTRACT

Biopreservatives are commonly used in food products to satisfy the increasing demand of consumers with increasing advancement in food and technology. The foods with chemical preservatives are now being neglected by the people and they prefer products which are generally recognized as safe (GRAS). Thus, as a result food industry is using naturally produced preservatives to increase the shelf life of product without any new technology. The most commonly used bio-preservatives are bacteriocins, essential oils, herbs and spices, vinegar, fermentation and sugar and salt. They exhibit growth inhibition of various microorganisms when added at different concentrations so as to preserve food products. These preservatives have been tested under laboratory conditions to know their apt use. This review provides an overview of the importance of bio-preservatives as per the increasing demand of consumers.

Keywords: *Bio-preservation; bacteriocins; essential oils; herbs and spices; vinegar; fermentation; salt and sugar.*

1. INTRODUCTION

The demand for production of food on a large scale is increasing tremendously due to increase in population and change in their food habits. Food products like meat and milk products are of perishable nature. Their shelf life can be increased by adding food preservatives which restrict the process of spoilage to some extent. The food industry is investigating more methods to preserve food products beyond the traditional methods which have been commonly used for thousands of years including curing, cooling, freezing, boiling, heating, sugaring, canning, pickling, fermentation and others. The new less invasive techniques are being developed such as use of high-pressure, pulse electric field, ultrasound, oscillating magnetic field, hurdle technology, hyperbaric pressure and UV treatment [1]. Biopreservatives are the new alternatives derived from natural sources to preserve and enhance the keeping quality of food and well suited for food application with increasing demand of consumers for chemical free foods.

To extend the shelf life of products by use of natural preservatives or controlled microbiota and/or antimicrobial compounds obtained from microbes is referred to as bio-preservation. The natural food preservatives preserve the food by lowering the pH value, altering water activity (a_w) and settling the redox potential of the product [2]. Most commonly selected organism for bio-preservation are lactic acid bacteria (LAB) and their metabolites. The functional foods with natural ingredients promoting health instead of synthetic additives have been intensively commercialized by the food industry [3,2]. During last decade, the food industries are looking for new natural bio-preservatives which can promote human health besides acting as food preservative [4]. The present communication provides an overview of the current scenario on bio-preservative compounds that are either actually being used industrially or under research and development stage.

2. BACTERIOCINS

Bacteriocins are secondary antimicrobial peptides that are ribosomally synthesized from the bacteria that exhibit antagonistic activity to some group of bacteria which can either be closely related or unrelated [5]. The hydroxyl (-OH) groups of bacteriocins interact with the bacterial cell membrane and causes disrupting

its structures and causing leakage of its components, which ultimately leads to their antimicrobial action [6]. Colicin was the first discovered bacteriocin in 1952 from *Escherichia coli* and showed bactericidal effects by binding to the inner membrane or other cytosolic targets causing permeabilization of cell membrane, inhibiting cell wall synthesis and inhibiting DNase or RNase activity on bacteria that are closely related [7]. The application of bacteriocins in various food products have been widely evaluated under different laboratory conditions. Bacteriocinogenic protective cultures can be used to increase the shelf life period of non-fermented foods by inhibiting spoilage and pathogenic bacteria [8]. Majority of the bacteriocins used commercially are produced by metabolic activity of Lactic Acid Bacteria like *Lactobacillus acidophilus* which have status of "Generally regarded as safe (GRAS)" because protease can degrade them [2]. Nisin and pediocin are the only commonly available bacteriocin in the food industry, produced by *L. lactis* and *Pediococcus acidilactici*, respectively. Bacteriocins produced from the bacterial strains exhibit the defensive advantage by competitive inhibition of the bacteria from other group for nutrition. In case of non-fermented food, bacteriocin producing bacterial strain used for their bio-preservation only when they do not have any undesirable effects on organoleptic properties of food. *L. brevis* is the most common spoilage organism in beer and is responsible for up to 52.5% contamination of beer. Nisin is a commercially used bacteriocin and is highly stable in the acidic solutions. It is incorporated in beer and is capable of killing 90% of the Gram-positive bacteria without affecting the fermentation activity of *Saccharomyces* yeast [9]. Lacticin 3147 is produced from the lactococcal strains, present in the Irish Kefir grain and is used in cheddar cheese manufacturing [10]. Nisin is being widely used in research as an inhibitor of heat shocked spore of *Clostridium* and *Bacillus* strains in canned foods. Minimum inhibitory concentration to nisin to prevent the outgrowth of spores ranges from 3 to >5,000 IU/ml [11]. They inhibit various food borne pathogens. *Actinomyces ruminicola* produced Actifensin having broad spectrum inhibitory activity against Gram-positive species including notable pathogens such as methicillin-resistant *Staphylococcus* and vancomycin-resistant *Enterococcus* [12]. Various food-borne and other pathogenic bacteria such as *Clostridium tyrobutyricum*, *C. sporogenes*, *Listeria monocytogenes* and *Staphylococcus aureus* can

be inhibited by Enterocins A and B isolated from *Enterococcus* sp. which also have a broad inhibitor spectrum against Gram-positive bacteria [13]. Properties of such bacteriocins and their usefulness against pathogenic organisms are detailed in Table 1.

3. ESSENTIAL OILS

Also commonly known as volatile odoriferous oils, are extracted from different plant materials like roots, barks, flowers, fruits etc and are aromatic liquids [14]. The approach of using essential oils as natural bio preservatives was intended for the purpose for preventing rancidity of fats and possible prevention of chronic degenerative disease. Essential oils (Eos) are using more often under the concept of hurdle technology with other food preservatives. Thus, it aids as “green technology” in food market. Earlier, these oils were primarily used as medicines and recently the food industry has adopted them as flavoring and coloring agents [15]. Colors of essential oils are due to the presence of indigenous pigments in plant material from where they are extracted. Because of their antimicrobial and antioxidant activities, they are widely and efficiently used in food products.

Steam distillation is commonly used process for the production of essential oils at commercial scale. They constitute low molecular weight organic compounds with vast anti-microbial activity. They have been broadly classified as Terpene Hydrocarbon and oxygenated compounds with main active compounds like terpenes, terpenoids, phenylpropenes etc. Essential oils are more prone to enzymatic and chemical reactions like oxidation, cyclization, isomerization or dehydrogenation when not provided with protective compartmentation, which further leads to quality loss. Oxidized terpenoids as well as some aged essential oils are known for organoleptic alterations, viscosity changes and even some skin sensitizing capacities [16,17,18,19]. Use of some of the essential oils as preservatives has been demonstrated by some of the researchers. Two herbal essential oils i.e., curry leaves and cloves were added at the rate of 0.10 ppm and 0.20 ppm respectively to enhance the storage stability of burfi without interfering with the sensory acceptability of the product [20].

The classification, mechanism of action and other applications of essential oils are summarized in Table 2.

4. HERBS AND SPICES

Plants have ability to synthesize various chemical compounds and these antimicrobial components enhance the defense mechanism of plant against natural infections. Many herbs and spices such as cloves (*Carophyllus aromaticus*), cinnamon, guar gum, mustard seed and garlic known to have antimicrobial properties. When added in food products, the antimicrobial properties associated with such herbs and spices offer them competency to prevent food from spoilage and exercise food safety by controlling growth of spoilage and pathogenic micro-organisms [21]. There are number of plants which are recognized as herbs or spices and are part of culinary practices followed in different countries to improve the sensory quality and shelf life of food products as well as owing to their nutritional, antimicrobial and health promoting properties. These properties are more intense in the extracts of spices rather than spices as such, due to the reason that spices release volatiles at a slower rate [22].

The main advantage of using herbs and spices in food as preservatives is due to their ‘generally recognized as safe’ (GRAS) status because their use in human diet is time tested. Further they are usually free from chemical residues and therefore are potential alternatives to chemical additives [23]. Herbs and spices contribute to the total dietary phenolic intake as they are the sources of polyphenols. The herbs and spices have great antioxidant property and consumption of herbs like cloves and garlic has positive effect on human health and they also have anti carcinogenic effect. The mechanism of action for herbs and spices has not been completely understood irrespective of the clear expression of antimicrobial activity. Butylated hydroxyanisole (BHA) prevents the auto oxidation of lipids and is seen to inhibit growth of various Gram negative as well as Gram positive bacteria when added in different food products. In a study conducted by Shelef and Liang [24], BHA added at a concentration of 5000 ppm in strained chicken and 1000 ppm in cooked rice exhibit growth inhibition of vegetative cells of *Bacillus* spp.

5. VINEGAR

Vinegar is a liquid solution and one of the most typical pickling agents with 5%-10% acetic acid and it preserves food by altering water activity or pH. It provides flavor to the product and also vitamins. The pH 4.6 is a distinguishing and

Table 1. Classification and properties of bacteriocins

| Class | Bacteriocins | Source | Properties | Target food contaminants | Reference |
|------------------------------|--|---|--|--|------------------|
| Class I (Lantibiotics) | Nisin | <i>Lactobacillus lactis</i> <i>subsp. Lactis</i> | Heat stable at low pH (2), resistant to trypsin, elastase, pepsin, carboxypeptidase A and sensitive to α -chymotrypsin. | <i>Staphylococcus aureus</i> , <i>Bacillus cereus</i> , <i>Clostridium botulinum</i> . | [25] |
| Class II | Lacticin | <i>L. lactis</i> | Stable at neutral and acidic pH | <i>L.monocytogenes</i> , <i>S.aureus</i> and <i>B.subtilis</i> | [26] |
| | Pediocin PA-1 | <i>Pediococcus acidilactici</i> | Stable at pH 4-6, resistant to DNAses, RNAses, lipase, catalase, lysozyme and phospholipase C. | <i>L.monocytogenes</i> , <i>Pediococcus pentosaceus</i> , <i>Lactobacillus helveticus</i> | [23] |
| | Enterocin 1071 | <i>Enterococcus faecalis</i> BFE 1071 | Sensitive to treatment with proteolytic enzymes | <i>Enterococcus faecalis</i> , <i>L. monocytogenes</i> , <i>L. innocua</i> | [27] |
| | Enterocin EJ97 | <i>E.faecalis</i> EJ97 | Sensitive to proteolytic enzymes | <i>E. faecalis</i> , <i>Bacillus spp.</i> , <i>L. monocytogenes</i> , <i>Geobacillus stearothermophilus</i> , <i>S. aureus</i> | [28] |
| Class III (Bacteriolysin) | Lytic Bacteriocin (Enterolysin A) | <i>E. faecalis</i> | Causes cell damage by attacking peptidoglycan layer of cell wall in susceptible Gram-positive bacteria | <i>Enterococcus spp.</i> | [29] |
| | Non lytic heat labile Bacteriocin (Helveticin J, Dysgalacticin) | | Hamper glucose uptake and strave the bacteria, hence depleting energy reserve causing cell death | <i>Streptococcus pyogenes</i> | [30] |
| Class IV | Enterocin AS-48 | <i>E. faecalis subsp. liquefaciens</i> S-48 | Compatible with several chemical compounds like EDTA, lactic acid, per acetic acid, phosphoric acid, sodium hypochlorite, hydrocinnamic acid | <i>Bacillus spp.</i> , <i>Geobacillus stearothermophilus</i> , <i>S.aureus</i> , <i>L. monocytogenes</i> , <i>Brocothrix thermosphacta</i> | [31] |

Table 2. Classification and properties of essential oils

| Chemical Classification | Example (Plant Origin) | Model Organism and Minimum Inhibitory Concentration (MIC) | Mechanism of Action | Reference |
|------------------------------------|---------------------------------|---|--|---------------------|
| Terpenes | p-Cymene (Oregano and Thyme) | <i>Staphylococcus aureus</i> (1250 µg/ml), <i>Escherichia coli</i> (2500 µg/ml) | Reduce cell mortality Decrease membrane potential Decrease membrane melting temperature | [32,33,34] |
| | α-Terpinene (Oregano and Thyme) | <i>Staphylococcus aureus</i> (2500-34000 µg/ml), <i>Escherichia coli</i> (5000 µg/ml) | Decrease membrane melting temperature Decrease transition enthalpy Might disrupt the membrane of microorganism | [35,33] |
| | Limonene (Lyme) | <i>Aspergillus flavus</i> (560 µg/ml), <i>A. parasiticus</i> (1130 µg/ml), <i>E. coli</i> , <i>S. aureus</i> | Reduce aflatoxin production Extra and intracellular damage to cells | [36] |
| Terpenoid | Carvacrol (Oregano and Thyme) | <i>Candida</i> strains (75-100 µg/ml), <i>Bacillus cereus</i> (900 µg/ml), <i>Salmonella typhimurium</i> (150-250 µg/ml), <i>Listeria monocytogenes</i> (450-1500 µg/ml) | Transient Calcium ion surge Act on specific signaling pathway Dissipated pH gradient and membrane potential Permeabilize cell membrane and vesicles | [33,37,38,39,1] |
| | Menthol (peppermint) | <i>C. albicans</i> (2500 µg/ml), <i>B. cereus</i> (1250 µg/ml), <i>Klebsiella pneumoniae</i> (2500 µg/ml), <i>Pseudomonas aeruginosa</i> (2500 µg/ml), <i>Proteus vulgaris</i> (1250 µg/ml) | Disrupt membrane permeability, Alter intracellular components, Release cellular content | [40,41,42] |
| | Linalool (Basil and Citrus oil) | <i>C. albicans</i> (2145 µg/ml), <i>B. cereus</i> (1073 µg/ml), <i>C. jejuni</i> (515 µg/ml), <i>L. monocytogenes</i> (1000-2145 µg/ml), <i>S. Typhimurium</i> (1000 µg/ml) | Permeabilize cell membrane | [43,35,44,45,1] |
| Phenylpropene Phenolic Aldehyde | Vanillin (Vanilla) | <i>L. innocua</i> (5325 µg/ml), Yeasts and molds (456-1460 µg/ml), <i>Lactobacillus plantarum</i> (11411 µg/ml), <i>E. coli</i> (2282 µg/ml) | Dissipates potassium and pH gradients, Activates ATP production in some cells | [46,47] |
| | Cinnamaldehyde (cinnamon) | <i>B. cereus</i> (0.3 µg/ml) <i>B. thermosphacta</i> (0.84 µg/ml) <i>E. coli</i> (397-1322 µg/ml) <i>E. faecalis</i> (250 µg/ml) | Concentration dependent ATPase inhibition Inhibited histidine decarboxylase and cytokinesis Inhibiting cell wall synthesizing enzymes | [48,39,49,50,51,52] |

characteristic feature of the process which doesn't allow the bacteria to grow and proliferate. Vinegar can be synthesized by alcoholic fermentation or the acetic acid fermentation. Vinegar provides an acidic medium to the food for preservation and improves the shelf life of the food product. Park et al. [53] stored the blanched tea leaves for 4 days at 30°C in pickling solutions as mixture of soy sauce, water and vinegar in different concentrations. The different parameters like color, pH, hardness, antioxidant compound content, sensory evaluation, acidity, ABTS radical scavenging were determined during storage. Authors observed that the total acidity and pH increased consistently and thereby imparted preservative effect. Jang and coworkers [54] conducted a study on Korean seasoned beef to examine vinegar and sake as preservation hurdles and detect their effect of sensory quality and microbial stability. They found that the combination of vinegar and sake did not improve the sensory quality however; microbial stability was improved both at 8°C and 20°C. Acetic acid has been used for decontamination of carcass to increase shelf life. Laboratory studies showed significant reduction of *E. coli* on rib-eye steaks treated with an acetic acid dip [55,56] studied prevention of Anthracnose rot (*Colletotrichum coccodes*) in Tomato (*Lycopersicon esculentum*) by treatment with Vinegar (VIN), Chlorine (CHL), absolute ethyl alcohol (AEA) and origanum oil (ORI) when stored at 12°C with 95% relative humidity. After treatment with AEA and VIN, the growth of mycelium was accelerated and exposure to pure VIN-, ORI vapors and CHL-decreased germination of spore's in vitro up to 92%.

6. FERMENTATION

Fermentation process produces numbers of beneficial products with the bacteria which helps in reduction of food spoilage and renders the food free from pathogenic microorganisms and metabolites [57]. The organisms used to serve this purpose mostly belong to the group of lactic acid bacteria (LAB). The metabolic compounds of these bacteria, such as organic acids are capable to exert antimicrobial properties as well as imparts unique flavor and texture to the food products [58]. There are certain advantages of fermentation carried out by LAB over other methods of food preservation. These include comparatively more availability of nutrients and ease in carrying out the process which require almost nil to little energy. Tarhana, a Turkish

fermented cereal food using baker's yeast and yoghurt bacteria as culture was studied to detect compositional changes in the organic acids developed during the fermentation phase of 3 days and subsequently stored for a period of 6 months. In a fermentation period of 3 days, lactic acid, titrable acidity, propionic acid and pyruvic acid increased from 13.58 to 20.26 g/kg, 26.50 to 41.4 g/kg, 2.44 to 7.58 g/kg and 0.16 to 0.58 g/kg respectively and citric acid content reduced from 6.39 to 3.58 g/kg [59]. Gotcheva et al. [60] monitored the fermentation of traditional Bulgarian beverage boza, a cereal based fermentation product prepared from both flour and whole wheat grains and the impact on product quality by raw material used for preparation. Yeast and Lactobacilli were the major organisms responsible for fermentation of boza and biochemical and physical changes were observed during initial 48 hour of fermentation. Glucose content was increased while viscosity, dry matter, pH and free amino nitrogen content were decreased. Two Nigerian fermented food products *Fufu* and *Ogi* from cassava and maize respectively were investigated for changes in pH, titrable acidity and temperature during fermentation by hetero fermentative *Lactobacillus* and *Leuconostoc*. pH decreased rapidly from 5.6 to 3.7 and 5.9 to 3.8 in *fufu* and *ogi* respectively and increase in temperature from 25°C to 31°C and 26°C to 30°C in *ogi* and *fufu* respectively [61]. A study conducted to stabilize meat cubes and minced meat with salt and glucose by process of fermentation. They used culture of *Lactobacillus casei*, *Lactobacillus plantarum* and *Lactococci lactis*. In an incubation of 24-36 hour at 37°C and 30-42 hour at 30°C, a significant reduction in *Escherichia coli*, *Salmonella* spp, coliforms and *Staphylococcus aureus* with a pH value of 4.0-4.2 was achieved [62]. *Rhizopus oligosporus* was used to ferment oat tempe and whole grain barley to investigate the effects of different pretreatment like rolling, moistening, pearling, autoclaving and soaking on mineral and phytate content. Phytate content in both oat and barley was decreased by 74% and 89% respectively by the most effective pearling process [63]. Fermented fish paste was prepared by fermenting at a room temperature for 8 and 32 days with addition of 2% NaCl from dried anchovy fish. Water content and pH values were increased during fermentation. Fatty acids except DHA, EPA and stearic acid decreased [64]. Microbial fermentation leads to some extremely complex interactions between bacterial species and the food matrix they are fermenting, this

represents an area with potential well beyond the extension of shelf life [65].

7. SUGAR AND SALT

Both salt and sugar exert food preservation effect by same mode. When added to the food product, it absorbs water and thereby inhibits the growth of microorganisms by restricting the water availability [66]. Sugar is added with salt to preserve fish and meat. It can be mixed with salt to form a dry mixture covering food or can be dissolved in liquid to make a brine to surround the food. Salt in water produces an isotonic condition for non-marine organism at concentrations of 0.85-0.90%. When salt is used in high concentrations to fresh meat, it undergoes plasmolysis which results in drying of meat and leads to death of microbial cells. The non-marine organism can be easily inhibited by <20% NaCl. Salt and sugar inhibit organisms at different concentrations. Sucrose needs about six times more concentration than salt to exhibit same degree of inhibition [67].

The research work conducted to study impact of sucrose solution on strawberry for three months at 15 days interval and data showed that treatments strawberry fruit with sucrose solution, Sodium benzoate and Potassium sorbate were found superior during physiochemical and organoleptic [68]. A study was conducted to find out the survival of indicator and pathogenic microorganism in seven liquid sweeteners. The sweeteners were inoculated with various species of bacteria at the rate of 10^5 cells per gram. The microorganism showed a significant reduction in less than 3 days when the inoculated products with sweeteners held at normal holding temperature. The slowest rate of reduction was observed for *S. aureus* [69]. Different hypertonic sucrose solutions with or without NaCl were used to treat apple samples osmotically. The acceptability of products was reduced when 0.5% NaCl was added than that by addition of 0.5% sucrose [70]. Effects of addition of salt and sugar on the enzymatic activity and quality of grass carp fish (*Ctenopharyngodon idellus*) filets were evaluated. The samples were grouped as uncured (CK), dry cured with 1.0% salt (T1) and dry cured with 1.0% salt and 1.0% sugar (T2). Authors observed that all curing treatments decreased TVB-N accumulation and improved the level of IMP in grass carp as compared to uncured samples [71]. Salt is used in cheddar cheese to control the development of bitter flavor by inhibiting the proteolysis of β -casein in

presence of 10% NaCl and was significantly reduced by use of 5% NaCl. In presence of 5-10% NaCl, rate of proteolysis was maximum of α -casein [72]. The survival of *Escherichia coli*, *Salmonella typhimurium*, *Staphylococcus aureus*, *Listeria monocytogenes*, *E. coli O157:H7* and *Clostridium perfringens* in natural casings at different water activity (a_w) levels was studied by preservation with NaCl to determine antimicrobial properties. The casings were stored at different temperatures in different brines with dry salt. The effect of salt was well sufficient to minimize the contamination of bacteria except the spores of *Clostridium* below acceptable level at a_w 0.85 or decreased after a storage period of 30 days [73].

8. CONCLUSION

There are several traditional and newly methods of bio-preservation of food are available. The common methods are preservation by bacteriocins, use of essential oils, different herbs and spices, vinegar, fermentation process, use of sugar and salt etc. All these methods are effective in their own way.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Kim J, Marshall MR, Wei CI. Antibacterial activity of some essential oil components against five food borne pathogens. *J. Agri Food Chem.* 1995;43(11):2839-2845.
2. Sharif ZIM, Mustapha FA, Jai J, Yusof NM, Zaki NAM. Review on methods of preservation and natural preservatives for extending the food longevity. *Chem Eng Res Bull.* 2017;19:145-153.
3. Carocho M, Barreiro M, Morales P, Ferreria ICFR. Adding molecules to food, pros and cons: A review on synthetic and natural food additives. *Compr Rev Food Sci F.* 2014;13:377-399.
4. Burt S. Essential oils: their antibacterial properties and potential applications in foods– A review. *Int J Food Microbiol.* 2004;94(3):223-253.
5. Johnson MEM, Jung YG, Jin YY, Jayabalan R, Yang SH, Suh JW. Bacteriocins as food preservatives: Challenges and emerging horizons. *Crit Rev Food Sci Nutr.*; 2017.

6. Quinto EJ, Caro I, Villalobos-Delgado LH, Mateo J, De-Mateo-Silleras B, Redondo-Del-Río MP. Food safety through natural antimicrobials. *Antibiotics*. 2019;8(4):208.
7. García-Bayona L, Guo SM, Laub TM. Contact-dependent killing by *Caulobacter crescentus* via cell surface-associated, glycine zipper proteins. *eLife*. 2017;6:e24869.
8. Gálvez A, Abriouel H, López RL, Omar NB. Bacteriocin-based strategies for food biopreservation. *International Journal of Food Microbiology*. 2007;120(1-2):51–70.
9. Müller-Auffermann K, Grijalva F, Jacob F, Hutzler M. Nisin and its usage in breweries: A review and discussion. *J Inst Brew*. 2015;121(3):309-319.
10. Ryan MP, Rea M, Hill C, Ross RP. An application in Cheddar cheese manufacture for a strain of *Lactococcus lactis* producing a novel broad-spectrum bacteriocin, lacticin 3147. *Appl Environ Microbiol*. 1996;62:612-9.
11. Hurst A. Nisin. *Adv Appl Microbiol*. 1981;27:85-123.
12. Sugrue I, O'Connor PM, Hill C, Stanton C, Ross RP. Actinomyces produce defensin-like bacteriocins (actifensins) with a highly degenerate structure and broad antimicrobial activity. *Journal of Bacteriology*; 2019.
13. Kubašová I, Diep DB, Ovchinnikov KV, Lauková A, Stropflová V. Bacteriocin production and distribution of bacteriocin-encoding genes in enterococci from dogs. *International Journal of Antimicrobial Agents*; 2019.
14. Tongnuanchan P, Benjakul S. Essential oils: Extraction, bioactivities, and their uses for food preservation. *J Food Sci*. 2014;79(7):1231-1249.
15. Hyldgaard M, Mygind T, Meyer RL. Essential oils in food preservation: Mode of action, synergies, and interactions with food matrix components. *Front Microbiol*. 2012;3(12):1-24.
16. Brared-Christensson J, Forsstrom P, Wennberg AM, Karlberg AT, Matura M. Air oxidation increases skin irritation from fragrance terpenes. *Contact Derm*. 2009;60:32–40.
17. Hagvall L, Skold M, Brared-Christensson J, Borje A, Karlberg AT. Lavender oil lacks natural protection against autoxidation, forming strong contact allergens on air exposure. *Contact Derm*. 2008;59:143–50.
18. Skold M, Hagvall L, Karlberg AT. Autoxidation of linalyl acetate, the main component of lavender oil, creates potent contact allergens. *Contact Derm*. 2008;58:9–14.
19. Woeber K, Krombach M. Zur Frage der Sensibilisierung durch atherische O le. *Berufsdermatosen*. 1969;17:320–326.
20. Badola R, Panjagari NR, Singh RRB, Singh AK, Prasad WG. Effect of clove bud and curry leaf essential oils on the antioxidative and anti-microbial activity of burfi, a milk-based confection. *J Food Sci Tech*. 2018;55(12):4802-4810.
21. Jack RW, Tagg JR, Ray B. Bacteriocins of gram positive bacteria. *Microbiol Rev*. 1995;59(2):171-200.
22. Shelef LA, Naglik OA, Bogen DW. Sensitivity of some common food borne bacteria to spices sage, rosemary and allspice. *J Food Sci*. 1980;45:7-12.
23. Rodriguez I, Guevara E. Dry matter production and nutritive value of the shrub legume *Cratylia argentea* in the south of Anzoategui State, Venezuela. *Revolucion Cientifica*. 2002;12(2):589-594.
24. Shelef LA, Liang P. Antibacterial effects of Butylated hydroxyanisole (BHA) against *Bacillus* species. *J Food Sci*. 1982;47:796-799.
25. Meghrous J, Lacroix C, Simard RE. The effects on vegetative cells and spores of three bacteriocins from lactic acid bacteria. *Food Microbiol*. 1999;16(2):105–114.
26. Ryan MP, Flynn J, Hill C, Ross RP, Meaney WJ. The natural food grade inhibitor, Lacticin 3147, reduced the incidence of mastitis after experimental challenge with *Streptococcus dysgalactiae* in nonlactating dairy cows. *J Dairy Sci*. 1999;82(10):2108–2114.
27. Balla E, Dicks LMT, DuToit M, Van Der Merwe MJ, Holzapfel WH. Characterization and cloning of the genes encoding enterocin 1071A and enterocin 1071B, two antimicrobial peptides produced by *Enterococcus faecalis* BFE 1071. *Appl Environ Microbiol*. 2000;66:1298-1304.
28. Galvez A, Valdivia E, Abriouel H, Camafeita E, Mendez E, Martinez-Bueno M, Maqueda M. Isolation and characterization of enterocin EJ97, a bacteriocin produced by *Enterococcus*

- faecalis* EJ97. Arch Microbiol. 1998;171(1): 59–65.
29. Nilsen T, Nes IF, Holo H. Enterolysin A, a cell wall-degrading bacteriocin from *Enterococcus faecalis* LMG 2333. Appl Environ Microbiol. 2003;69(5):2975–2984.
 30. Heng NCK, Swe PM, Ting YT, Dufour M, Baird HJ, Ragland NL, Burtenshaw GA, Jack RW, Tagg JR. The large antimicrobial proteins (bacteriocins) of streptococci. Int Congr. 2006;1289:351–354.
 31. Cobo Molinos A, Abriouel H, Lopez RL, Valdivia E, Omar NB, Galvez A. Combined physio-chemical treatments based on enterocin AS-48 for inactivation of gram-negative bacteria in soybean sprouts. Food Chem Toxicol. 2008;46(8):2912–21.
 32. Burt SA, Van Der Zee R, Koets AP, De Graaff AM, Van Knapen F, Gaastra W, Haagsman HP, Veldhuizen EJA. Carvacrol induces heat shock protein 60 and inhibits synthesis of flagellin in *Escherichia coli* O157:H7. Appl Environ Microbiol. 2007;73: 4484–4490.
 33. Cristani M, D'Arrigo M, Mandalari G, Castelli F, Sarpietro MG, Micieli D, Venuti V, Bisignano G, Saija A, Trombetta D. Interaction of four monoterpenes contained in essential oils with model membranes implications for their antibacterial activity. J Agri Food Chem. 2007;55:6300–6308.
 34. Ultee A, Bennik MHJ, Moezelaar R. The phenolic hydroxyl group of carvacrol is essential for action against the food-borne pathogen *Bacillus cereus*. Applied and Environment Microbiology. 2002;68:1561–1568.
 35. Carson CF, Riley TV. Antimicrobial activity of the major components of the essential oil of *Melaleuca alternifolia*. J Appl Bacteriol. 1995;78:264–269.
 36. Rammanee K, Hongpattarakere T. Effects of tropical citrus essential oils on growth, aflatoxin production, and ultrastructure alterations of *Aspergillus flavus* and *Aspergillus parasiticus*. Food Bio-process Technol. 2011;4:1050–1059.
 37. Gill AO, Holley RA. Disruption of *Escherichia coli*, *Listeria monocytogenes* and *Lactobacillus sakei* cellular membranes by plant oil aromatics. Int J Food Microbiol. 2006a;108:1–9.
 38. Gill AO, Holley RA. Inhibition of membrane bound ATPase's of *Escherichia coli* and *Listeria monocytogenes* by plant oil aromatics. Int J Food Microbiol. 2006b;111:170–174.
 39. Helander IM, Alakomi HL, Latva-Kala K, Mattila-Sandholm T, Pol I, Smid EJ, Gorris LGM, Von Wright A. Characterization of the action of selected essential oil components on gram-negative bacteria. J Agri Food Chem. 1998;46:3590–3595.
 40. Bassole IHN, Lamien-Meda A, Bay-ala B, Tirogo S, Franz C, Novak J, Nebie RC, Dicko MH. Composition and antimicrobial activities of *Lippia multi-flora* Moldenke, *Mentha x piperita* L. and *Ocimum basilicum* L. essential oils and their major monoterpene alcohols alone and in combination. Molecules. 2010;15:7825–7839.
 41. Iscan G, Kirimer N, Kurkcuoglu M, Baser KHC, Demirci F. Antimicrobial screening of *Mentha piperita* essential oils. J Agri Food Chem. 2002;50:3943–3946.
 42. Trombetta D, Castelli F, Sarpietro MG, Venuti V, Cristani M, Daniele C, Saija A, Mazzanti G, Bisignano G. Mechanisms of antibacterial action of three monoterpenes. Antimicrob Agents Chemother. 2005;49: 2474–2478.
 43. Bagamboula CF, Uyttendaele M, Debevere J. Inhibitory effect of thyme and basil essential oils, carvacrol, thymol, estragol, linalool and p-cymene towards *Shigella son-nei* and *S. flexneri*. Food Microbiol. 2004;21:33–42.
 44. Fisher K, Phillips C. Potential antimicrobial uses of essential oils in food: Is citrus the answer? Trends Food Sci Technol. 2008;19:156–164.
 45. Fisher K, Phillips CA. The effect of lemon, orange and bergamot essential oils and their components on the survival of *Campylobacter jejuni*, *Escherichia coli* O157, *Listeria monocytogenes*, *Bacillus cereus* and *Staphylococcus aureus* in vitro and in food systems. Journal of Appl Microbiol. 2006;101:1232–1240.
 46. Fitzgerald DJ, Stratford M, Gasson MJ, Narbad A. Structure-function analysis of the vanillin molecule and its antifungal properties. J Agri Food Chem. 2005;53: 1769–1775.
 47. Fitzgerald DJ, Stratford M, Narbad A. Analysis of the inhibition of food spoilage yeasts by vanillin. Int J Food Microbiol. 2003;86:113–122.
 48. Bang KH, Lee DW, Park HM, Rhee YH. Inhibition of fungal cell wall synthesizing

- enzymes by trans-cinnamaldehyde. *Biosci Biotech Biochem.* 2000;64:1061–1063.
49. Kwon JA, Yu CB, Park HD. Bacteriocidal effects and inhibition of cell separation of cinnamic aldehyde on *Bacillus cereus*. *Lett Appl Microbiol.* 2003;37:61–65.
 50. Wendakoon CN, Morihiko S. Inhibition of amino acid decarboxylase activity of *Enterobacter aerogenes* by active components in spices. *J Food Prot.* 1995;58:280–283.
 51. Yamazaki K, Yamamoto T, Kawai Y, Inoue N. Enhancement of antilisterial activity of essential oil constituents by nisin and diglycerol fatty acid ester. *Food Microbiol.* 2004;21:283–289.
 52. Zemek J, Valent M, Podova M, Kosikova B, Joniak D. Antimicrobial properties of aromatic compounds of plant origin. *Folia Microbiol.* 1987;32:421–425.
 53. Park BR, Park JJ, Hwang IG, Han HM, Shin M, Shin DS, Yoo SM. Quality and antioxidant activity characteristics during storage of tea leaf pickles with different vinegar contents. *Korean J Food Cook Sci.* 2014;30(4):402-411.
 54. Jang JD, Seo GH, Lyu ES, Yam KL, Lee DS. Hurdle effect of vinegar and sake on Korean seasoned beef preserved by sous vide packaging. *Food Cont.* 2006;17:171–175.
 55. Kotula K, Thelappurate R. Microbiological and sensory attributes of retail cuts of beef treated with acetic and lactic acid solutions. *J Food Prot.* 1994;57:665–670.
 56. Tzortzakis NG. Ethanol, vinegar and *Origanum vulgare* oil vapor suppress the development of anthracnose rot in tomato fruit. *Int J Food Microbiol.* 2010;142(1-2): 14–18.
 57. Ganguly S. Basic principles for effective food preservation: a review. *Int J Pure Appl Biosci.* 2013;1(6):84–85.
 58. Lucera A, Costa C, Conte A, Del Nobile MA. Food applications of natural antimicrobial compounds. *Front Microbiol.* 2012;3:287.
 59. Erbas M, Kemal Uslu M, Ozgun Erbas M, Certel M. Effects of fermentation and storage on the organic and fatty acid contents of tarhana, a Turkish fermented cereal food. *J Food Compos Anal.* 2006;19(4):294-301.
 60. Gotcheva V, Pandiella SS, Angelov A, Roshkova Z, Webb C. Monitoring the fermentation of the traditional Bulgarian beverage boza. *Int J Food Sci Technol.* 2001;36(2):129-134.
 61. Oyedeji O, Ogunbanwo ST, Onilude AA. Predominant lactic acid bacteria involved in the traditional fermentation of Fufu and Ogi, Two Nigerian fermented food products. *Food Nutr Sci.* 2013;4:40-46.
 62. Sakhare PZ, Narasimha Rao D. Microbial profiles during lactic fermentation of meat by combined starter cultures at high temperatures. *Food Control.* 2003;14(1):1-5.
 63. Eklund-Jonsson C, Sandberg AS, Alminger ML. Reduction of phytate content while preserving minerals during whole grain cereal tempe fermentation. *J Cereal sci.* 2006;44(2):154-160.
 64. Anggo AD, Ma Ruf WF, Swastawati F, Rianingsih L. Changes of amino and fatty acids in anchovy (*Stolephorus* Sp) fermented fish paste with different fermentation periods. *Procedia Environ Sci.* 2015;23:58-63.
 65. Hill D, Sugrue I, Arendt E, Hill C, Stanton C, Ross RP. Recent advances in microbial fermentation for dairy and health. *F1000Res.* 2017;6:751.
 66. Dwivedi S, Prajapati P, Vyad N, Malviya S, Kharia A. A review on food preservation: Methods, harmful effects and better alternatives. *Asian J Pharma Pharmacol.* 2017;3(6):193-199.
 67. Jey JM. *Modern food microbiology.* 5th Edn, ASPN Publishers Inc. Maryland. USA; 1998.
 68. Khan A, Shamrez B, Litaf U, Zeb A, Rehman Z. Effect of sucrose solution and chemical preservatives on overall quality of strawberry fruit. *J Food Proces Technol.* 2015;6:413.
 69. Niroomand F, Sperber WH, Lewandowski VJ, Hobbs LJ. Fate of bacterial pathogens and indicator organism in liquid sweeteners. *J Food Prot.* 1998;61:295-299.
 70. Sacchetti G, Gianotti A, Rosa MD. Sucrose-salt combined effects on mass transfer kinetics and product acceptability. Study on apple osmotic treatments. *J Food Eng.* 2001;49:163-173.
 71. Quin N, Zhang Y, Luo Y. Effects of adding salt and sugar on the quality and IMP-related enzyme activity of Grass Carp

- (*Ctebopharyngdon idellus*) fillets during 0°C. J Food Proces Preser. 2016;41(2): e12844.
DOI: 10.1111/jfpp.12844
72. Fox PF, Walley BF. Influence of sodium chloride on the proteolysis of casein by rennet and by pepsin. J Dairy Res. 1971;38(2):165-170.
73. Wijnker JJ, Koop G, Lipman LJA. Antimicrobial properties of salt (NaCl) used for the preservation of natural casings. Food Microbiol. 2006;23(7):657-662.

© 2019 Vaishali et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/53215>